

Original Article

Evaluation of *ABCB1* Gene Promoter Methylation in Patients with Ulcerative Colitis

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Abstract

Background: The pathogenesis of inflammatory bowel disease may be associated with the disruption in interactions between the immune system and gut flora. Epigenetic mechanisms especially, DNA methylation appear to be significant regarding the interaction between the environment and genome. *ABCB1* is the encoding gene for multi-drug resistance protein 1 (MDR1) (P-glycoprotein), which is an important transmembrane protein responsible for the efflux of cellular molecules from the intestinal wall to the lumen.

Method: In this study, we compared the methylation status of the promoter of *ABCB1* in rectal mucosa of patients with ulcerative colitis (UC) and healthy controls by using the bisulfite conversion system and real-time quantitative multiplex methylation-specific PCR (QM-MSP).

Results: We demonstrated that the mucosal specimen of 26 UC patients had significantly higher levels of promoter methylation in comparison to 26 controls.

Conclusion: As the first investigation of Iranian patients with UC, we showed that patients had higher levels of *ABCB1* promoter methylation in their inflammatory rectal mucosa compared to controls. However, this altered state of methylation did not associate with the characteristics of the patients such as age and sex. Our findings are a basis for further studies on concurrent assessment of promoter methylation and expression of *ABCB1* in UC.

Keywords: *ABCB1*; Epigenetics; Gene Methylation; MDR1; Ulcerative Colitis

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Introduction

Inflammatory bowel disease (IBD) is a chronic heterogeneous gastrointestinal inflammation that has two subtypes including Crohn's disease (CD) and ulcerative colitis (UC) (1). Over the past years, IBD has become more prevalent in developing countries. Notwithstanding, IBD incidence has barely changed in developed countries such as in European and North American countries (2, 3). In contrast to Europe and the US, IBD has been an infrequent disorder in Asia (4). However, the incidence and prevalence of IBD is now growing in Asia as well (5).

To date, related studies have suggested that the triggering incident in the pathogenesis of IBD may be the disruption in interactions between the immune system and flora within the intestine, though, the accurate etiology of this disease is still ambiguous (1, 6). Several studies have demonstrated the onset and progression of IBD are associated with approximately 32 predisposing loci (7-10). It is estimated that merely 20 percent of genetic susceptibility is due to these predisposing loci (11, 12) implying that the pathogenesis of IBD is probably correlated with other distinct changes such as epigenetic alterations (13). More specifically, there are several models of epigenetic alterations, from which DNA methylation is one of the most investigated. For instance, it has been reported that P14ARF, E-cadherin, and estrogen receptor (ER) genes undergo inappropriate DNA methylation in UC individuals (14). Moreover, an increasing number of studies support this notion that epigenetic changes are of the essence in the modulation of gene expression in patients with IBD (14, 15).

ABCB1 is the encoding gene for MDR1 protein (P-glycoprotein), which is an important transmembrane protein responsible for the efflux of cellular molecules from the intestinal wall to the lumen. It has the highest expression in the apical epithelium of the intestine (16). It is a member of the gene family encoding ATP-binding cassette (ABC) transporters and showed altered methylation and expression in different types of cancer (17). Patients with IBD are also at increased risk of developing CRC (18). Recently, it has been demonstrated that gastrointestinal inflammatory diseases e.g. IBD, are associated with alterations in ABCB1 expression (19). In this regard, a bulk

of animal studies indicated that inflammatory conditions in the intestine are correlated with decreased expression and activity of ABCB1 (20-22). Furthermore, it was revealed there is an association between ABCB1 single nucleotide polymorphisms (SNPs) and IBD pathogenesis (23-25). Overall, data shows a decreased level of ABCB1 expression and/or function in inflammatory conditions of the intestine. As it is documented that DNA methylation is an essential epigenetic factor involved in altered protein expression and it gets exacerbated in chronic inflammatory conditions; in this study, we assessed the level of DNA methylation in the ABCB1 gene in the inflammatory mucosa of UC patients.

Methods

Participants

The participants of the study consisted of 26 UC patients along with a group of 26 healthy controls (HCs) enrolled at the gastroenterology clinics of Shahid Sadoughi Hospital in Yazd, Iran, between May 2016 and July 2017. The diagnosis of UC was made based on clinical impression and confirmed pathology. Individuals with normal biopsy, which has been taken to rule out microscopic colitis, were included as healthy controls. Besides, written informed consents were obtained from all patients and healthy controls before inclusion in the study.

DNA isolation and bisulfite modification

Rectal mucosal specimens were taken from the patients and HCs. Specimens were taken during a colonoscopic biopsy and preserved at -80°C until use.

The genomic DNA was extracted from the specimens by using the High Pure PCR Template Preparation Kit (Roche). DNA bisulfite modification was performed by using the MethylEdge™ Bisulfite Conversion System (Promega Co) Based on the protocol, 20µl of each genomic DNA sample with optimal concentration (200–500ng) and high quality (optical density: 1.8-2) were applied. Through DNA bisulfite modification, the unmethylated cytosine residues are converted to uracil, which in turn will be converted to thiamine following the PCR. On the contrary, the methylated cytosine (5-methylcytosine) of the genomic DNA remains intact.

Quantitative Multiplex Methylation-Specific PCR

The promoter DNA methylation of the ABCB1 gene was assessed by the real-time quantitative multiplex methylation-specific PCR (QM-MSP) method. First, bisulfite-specific primers (BSP) or external primers were used for amplification of the 271 bp amplicon from the converted genomic DNA. The forward and reverse external primers are listed in **Table 1**. In each reaction well, 1µl of the converted DNA along with 19 µl of other reaction materials was applied. The sequence of reactions included one cycle of 95°C for 5min, then 30 cycles of 94°C for 30s, 58°C for 30s, 72°C for 30s, and finally 72°C for 5min.

Then the real-time PCR was performed on the product of the previous PCR to amplify a 75 bp amplicon by using methylation-specific primers (MSP) or internal primers (**Table 1**). The UCSC database was utilized to examine the methylation status of CpG islands in the promoter of ABCB1 and the Meth Blast tool was utilized for primer blasting. In each reaction well, 1µl of the BSP product as a template along with 5µl SYBR® Green Master Mix, 3 µl DDW, and 1µl of the MSP were applied. The sequence of reactions included one cycle of 95°C for 5min, then 30 cycles of 94°C for 30s, 60°C for 30s, 72°C for 30s, and finally 72°C for 5min.

The $\Delta\Delta Ct$ method was used to calculate the

Table 1. External (bisulfite-specific) and internal (methylation-specific) primers used in conventional and real time PCR

Primer Name	Primer Sequence (5' → 3')
EF	TTGTGGTGAGGTTGATTGGTT
ER	CCCAACTTTACGTACCCCTACC
IF	CGTGGGTTGAGTATAGTCGTTT
IR	ACGTACCCCTACCTCGCG
EF, External Forward; ER, External Reverse; IF, Internal Forward; IR, Internal Reverse	

unmethylated DNA level in each sample. The BSP was used to normalize data and a converted methylated human control DNA (Promega, N1221) was used as a reference. The following formula, which is used for relative gene expression (26, 27), was applied:

$$\text{Unmethylated DNA Level} = 2^{-\Delta\Delta Ct}$$

$$\Delta\Delta Ct = \Delta Ct \text{ sample} - \Delta Ct \text{ reference}$$

$$\Delta Ct \text{ sample} = CtMSP - CtBSP$$

$$\Delta Ct \text{ reference} = CtMSP - CtBSP$$

Statistical analysis

Statistical analyses were performed by using SPSS 20. The Chi-squared test was used for the comparison of sex between the two study groups. Student t-test was used to compare age between UC patients and HCs. The Mann-Whitney U test was used to compare the levels of promoter methylation between the patients and HCs. A *P*-value

< 0.05 was considered statistically significant.

Results

The promoter methylation of the ABCB1 gene was assessed in the rectal inflammatory mucosal samples of 26 patients with UC and 26 paired HCs. The characteristics of patients and HCs are illustrated in **Table 2**. The patient and control group were matched based on age. We demonstrated that the mucosal specimen of the UC patient had significantly higher levels of promoter methylation compared to HCs [median (IQR) for relative unmethylated level, 3.7×10^{-6} (2.4×10^{-6}) vs 4.2×10^{-6} (2.6×10^{-6}), *P*-value = 0.010]. However, there was no association between age and the level of promoter methylation in UC patients (*P*-value=0.444). Furthermore, sex was not associated with promoter methylation in UC patients (*P*-value=0.778).

Table 2. Clinical and methylation characteristics of UC patients and healthy controls

Variables	UC patients (n=26)	HCs (n=26)	<i>P</i> -value
Age [#]	34.4 ± 13.5	37 ± 4.5	0.554 ^a
Sex (male %)	19 (73%)	12 (48%)	0.089 ^b
Unmethylated ABCB1 Promoter (median [IQR])	3.7×10^{-6} [2.4×10^{-6}]	4.2×10^{-6} [2.6×10^{-6}]	0.010 ^c

[#] for UC n=17 and HC n=3, ^a student t-test, ^b Chi-squared test, ^c Mann-Whitney U test

Discussion

Epigenetic mechanisms appear to be of significant value regarding the interaction between the environment and genome (28). Further, than germline DNA alterations, epigenetic alterations, for instance, DNA methylation and histone modifications can influence gene function leading to disease-associated manifestations (29). In another words, epigenetic alterations are potential role players in disease development when the susceptible genetic loci are not responsible. In line with other autoimmune diseases, the exact pathological mechanisms behind IBD, and specifically UC are unknown. However, it has been speculated that the interactions between the host and the environment are involved in the pathogenesis of UC (1, 30). The significant factors in these interactions include the microbiota, immune responses, genetic susceptibility of the host, and particular environmental factors such as smoking, breastfeeding, drugs, and dietary products (1, 31, 32). Growing evidence revealed the hypermethylation of many gene promoters in UC. In this regard, it was demonstrated the incorporation of the 3H-methyl group into DNA was enhanced in patients with UC compared to HCs. The pattern was also observed in patients with histologically active compared to inactive disease (33). Moreover, hypermethylation of gene promoters of *ABCB1*, *E-cadherin*, *CDH1*, *GDNF*, and *p16* were reported in UC patients (15, 34). Overall, DNA methylation has been considered a player in the pathogenesis of UC, which has especially been reported in specimens from inflamed tissue. Also, DNA methylation in UC is associated with several determinants including disease severity, duration, active inflammation, and dysplasia (15, 35-37).

The *ABCB1* gene is located on chromosome 7 at q21.1 (38) and its altered methylation status has been reported in many types of disorders including malignancy and autoimmunity (17, 39). It is considered one of the susceptibility loci to IBD in genome-wide analysis in a UK cohort (40) and was further confirmed by genome scan meta-analysis (41). Lots of polymorphism studies have reported the association between *ABCB1* and UC. Independent studies in Europe demonstrated that a C3435T polymorphism in exon 26 of *ABCB1* is associated with susceptibility to UC

but not to CD (25, 42). A Japanese study reported an association between the *ABCB1* C3435T polymorphism and late-onset UC (43). *ABCB1* C1236T polymorphisms are reported to be associated with susceptibility to UC (44). Moreover, a number of studies showed decreased expression of *ABCB1* in UC patients (45, 46).

Considering the role of *ABCB1* protein as an energy-dependent efflux pump to transport different types of substances such as ions and peptides (47), through the efflux of toxins, it prevents mitochondrial damage. Decreased expression of *ABCB1* associated with mitochondrial damage (48). A previous study also showed that mice with polymorphism in the mitochondrial gene which generates high ATP are protected against colitis (49). The mitochondria are a key player in energy production. It has been suggested that there is an important connection between colonic epithelial cells' energy deficiency through mitochondrial damage and the pathophysiology of IBD (50). Based on the significant role of *ABCB1* in the pathogenesis of UC, we investigated the methylation status of *ABCB1* promoter in patients with UC and HCs in this experiment. In line with previous studies, we demonstrated that patients with UC have higher levels of methylation in the promoter of the *ABCB1* gene.

Conclusion

This is the first investigation into the methylation status in the promoter of the *ABCB1* gene in Iranian patients with UC. We showed that patients with UC have higher levels of *ABCB1* promoter methylation in their rectal inflammatory mucosa. However, this altered state of methylation did not associate with characteristics of the patients such as age and sex. Our findings highlight a need for further studies on concurrent assessment of promoter methylation and expression of *ABCB1* in UC.

Ethics approval and consent to participate

The protocol of this study was approved by the Ethics Committee of Tehran University of Medical Sciences and written informed consent was obtained from all participants before inclusion in the study.

Conflict of interests

The authors have no conflict of interest.

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References

- Kaser A, Zeissig S, Blumberg RS. Inflammatory bowel disease. *Annu Rev Immunol*. 2010;28:573-621.
- Molodecky NA, Soon IS, Rabi DM, Ghali WA, Ferris M, Chernoff G, et al. Increasing incidence and prevalence of the inflammatory bowel diseases with time, based on systematic review. *Gastroenterology*. 2012;142(1):46-54 e42; quiz e30.
- Shanahan F, Bernstein CN. The evolving epidemiology of inflammatory bowel disease. *Curr Opin Gastroenterol*. 2009;25(4):301-5.
- Loftus EV, Jr. Clinical epidemiology of inflammatory bowel disease: Incidence, prevalence, and environmental influences. *Gastroenterology*. 2004;126(6):1504-17.
- Thia KT, Loftus EV, Jr., Sandborn WJ, Yang SK. An update on the epidemiology of inflammatory bowel disease in Asia. *Am J Gastroenterol*. 2008;103(12):3167-82.
- Jones PA, Takai D. The role of DNA methylation in mammalian epigenetics. *Science*. 2001;293(5532):1068-70.
- Duerr RH, Taylor KD, Brant SR, Rioux JD, Silverberg MS, Daly MJ, et al. A genome-wide association study identifies IL23R as an inflammatory bowel disease gene. *Science*. 2006;314(5804):1461-3.
- Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. *Nat Genet*. 2007;39(2):207-11.
- Rioux JD, Xavier RJ, Taylor KD, Silverberg MS, Goyette P, Huett A, et al. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. *Nat Genet*. 2007;39(5):596-604.
- Yamazaki K, McGovern D, Ragoussis J, Paolucci M, Butler H, Jewell D, et al. Single nucleotide polymorphisms in TNFSF15 confer susceptibility to Crohn's disease. *Hum Mol Genet*. 2005;14(22):3499-506.
- Barrett JC, Hansoul S, Nicolae DL, Cho JH, Duerr

RH, Rioux JD, et al. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat Genet*. 2008;40(8):955-62.

- Xavier RJ, Rioux JD. Genome-wide association studies: a new window into immune-mediated diseases. *Nat Rev Immunol*. 2008;8(8):631-43.
- Petronis A, Petroniene R. Epigenetics of inflammatory bowel disease. *Gut*. 2000;47(2):302-6.
- Maeda O, Ando T, Watanabe O, Ishiguro K, Ohmiya N, Niwa Y, et al. DNA hypermethylation in colorectal neoplasms and inflammatory bowel disease: a mini review. *Inflammopharmacology*. 2006;14(5-6):204-6.
- Tahara T, Shibata T, Nakamura M, Yamashita H, Yoshioka D, Okubo M, et al. Effect of MDR1 gene promoter methylation in patients with ulcerative colitis. *Int J Mol Med*. 2009;23(4):521-7.
- Thiebaut F, Tsuruo T, Hamada H, Gottesman MM, Pastan I, Willingham MC. Cellular localization of the multidrug-resistance gene product P-glycoprotein in normal human tissues. *Proc Natl Acad Sci U S A*. 1987;84(21):7735-8.
- Zappe K, Cichna-Markl M. Aberrant DNA Methylation of ABC Transporters in Cancer. *Cells*. 2020;9(10).
- Triantafyllidis JK, Nasioulas G, Kosmidis PA. Colorectal cancer and inflammatory bowel disease: epidemiology, risk factors, mechanisms of carcinogenesis and prevention strategies. *Anticancer Res*. 2009;29(7):2727-37.
- Ho GT, Moodie FM, Satsangi J. Multidrug resistance 1 gene (P-glycoprotein 170): an important determinant in gastrointestinal disease? *Gut*. 2003;52(5):759-66.
- Banner KH, Cattaneo C, Le Net JL, Popovic A, Collins D, Gale JD. Macroscopic, microscopic and biochemical characterisation of spontaneous colitis in a transgenic mouse, deficient in the multiple drug resistance 1a gene. *Br J Pharmacol*. 2004;143(5):590-8.
- Kalitsky-Szirtes J, Shayeganpour A, Brocks DR, Piquette-Miller M. Suppression of drug-metabolizing enzymes and efflux transporters in the intestine of endotoxin-treated rats. *Drug Metab Dispos*. 2004;32(1):20-7.
- Wilk JN, Bilsborough J, Viney JL. The *mdr1a*^{-/-} mouse model of spontaneous colitis: a relevant and appropriate animal model to study inflammatory bowel disease. *Immunol Res*. 2005;31(2):151-9.
- Brant SR, Panhuysen CI, Nicolae D, Reddy DM, Bonen DK, Karaliukas R, et al. MDR1 Ala893 polymorphism is associated with inflammatory bowel disease. *Am J Hum Genet*. 2003;73(6):1282-

- 92.
24. Hoffmeyer S, Burk O, von Richter O, Arnold HP, Brockmoller J, Johne A, et al. Functional polymorphisms of the human multidrug-resistance gene: multiple sequence variations and correlation of one allele with P-glycoprotein expression and activity in vivo. *Proc Natl Acad Sci U S A*. 2000;97(7):3473-8.
25. Schwab M, Schaeffeler E, Marx C, Fromm MF, Kaskas B, Metzler J, et al. Association between the C3435T MDR1 gene polymorphism and susceptibility for ulcerative colitis. *Gastroenterology*. 2003;124(1):26-33.
26. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C(T) method. *Nat Protoc*. 2008;3(6):1101-8.
27. Hussein MI, Kuroda A, Kaye AN, Nair I, Kandeel F, Ferreri K. Development of a quantitative methylation-specific polymerase chain reaction method for monitoring beta cell death in type 1 diabetes. *PLoS One*. 2012;7(10):e47942.
28. Bird AP, Wolffe AP. Methylation-induced repression--belts, braces, and chromatin. *Cell*. 1999;99(5):451-4.
29. Petronis A. Epigenetics as a unifying principle in the aetiology of complex traits and diseases. *Nature*. 2010;465(7299):721-7.
30. Yi JM, Kim TO. Epigenetic alterations in inflammatory bowel disease and cancer. *Intest Res*. 2015;13(2):112-21.
31. Jostins L, Ripke S, Weersma RK, Duerr RH, McGovern DP, Hui KY, et al. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature*. 2012;491(7422):119-24.
32. Kim JM. Antimicrobial proteins in intestine and inflammatory bowel diseases. *Intest Res*. 2014;12(1):20-33.
33. Gloria L, Cravo M, Pinto A, de Sousa LS, Chaves P, Leitao CN, et al. DNA hypomethylation and proliferative activity are increased in the rectal mucosa of patients with long-standing ulcerative colitis. *Cancer*. 1996;78(11):2300-6.
34. Hsieh CJ, Klump B, Holzmann K, Borchard F, Gregor M, Porschen R. Hypermethylation of the p16INK4a promoter in colectomy specimens of patients with long-standing and extensive ulcerative colitis. *Cancer Res*. 1998;58(17):3942-5.
35. Kuester D, Guenther T, Biesold S, Hartmann A, Bataille F, Ruemmele P, et al. Aberrant methylation of DAPK in long-standing ulcerative colitis and ulcerative colitis-associated carcinoma. *Pathol Res Pract*. 2010;206(9):616-24.
36. Saito S, Kato J, Hiraoka S, Horii J, Suzuki H, Higashi R, et al. DNA methylation of colon mucosa in ulcerative colitis patients: correlation with inflammatory status. *Inflamm Bowel Dis*. 2011;17(9):1955-65.
37. Azarschab P, Porschen R, Gregor M, Blin N, Holzmann K. Epigenetic control of the E-cadherin gene (CDH1) by CpG methylation in colectomy samples of patients with ulcerative colitis. *Genes Chromosomes Cancer*. 2002;35(2):121-6.
38. Callen DF, Baker E, Simmers RN, Seshadri R, Roninson IB. Localization of the human multiple drug resistance gene, MDR1, to 7q21.1. *Hum Genet*. 1987;77(2):142-4.
39. Moret-Tatay I, Cerrillo E, Saez-Gonzalez E, Hervas D, Iborra M, Sandoval J, et al. Identification of Epigenetic Methylation Signatures With Clinical Value in Crohn's Disease. *Clin Transl Gastroenterol*. 2019;10(10):e00083.
40. Satsangi J, Parkes M, Louis E, Hashimoto L, Kato N, Welsh K, et al. Two stage genome-wide search in inflammatory bowel disease provides evidence for susceptibility loci on chromosomes 3, 7 and 12. *Nat Genet*. 1996;14(2):199-202.
41. van Heel DA, Fisher SA, Kirby A, Daly MJ, Rioux JD, Lewis CM, et al. Inflammatory bowel disease susceptibility loci defined by genome scan meta-analysis of 1952 affected relative pairs. *Hum Mol Genet*. 2004;13(7):763-70.
42. Ho GT, Nimmo ER, Tenesa A, Fennell J, Drummond H, Mowat C, et al. Allelic variations of the multidrug resistance gene determine susceptibility and disease behavior in ulcerative colitis. *Gastroenterology*. 2005;128(2):288-96.
43. Osuga T, Sakaeda T, Nakamura T, Yamada T, Koyama T, Tamura T, et al. MDR1 C3435T polymorphism is predictive of later onset of ulcerative colitis in Japanese. *Biol Pharm Bull*. 2006;29(2):324-9.
44. Cao Y, Qu C, Chen Y, Li L, Wang X. Association of ABCB1 polymorphisms and ulcerative colitis susceptibility. *Int J Clin Exp Pathol*. 2015;8(1):943-7.
45. Englund G, Jacobson A, Rorsman F, Artursson P, Kindmark A, Ronnblom A. Efflux transporters in ulcerative colitis: decreased expression of BCRP (ABCG2) and Pgp (ABCB1). *Inflamm Bowel Dis*. 2007;13(3):291-7.
46. Yamamoto-Furusho JK, Villeda-Ramirez MA, Fonseca-Camarillo G, Sanchez-Munoz F, Dominguez-Lopez A, Barreto-Zuniga R, et al. High gene expression of MDR1 (ABCB1) is associated with medical treatment response and long-term remission in patients with ulcerative colitis. *Inflamm Bowel Dis*. 2010;16(4):541-2.
47. Gupta S, Gollapudi S. P-glycoprotein (MDR1 gene product) in cells of the immune system: its possible physiologic role and alteration in aging and human immunodeficiency virus-1 (HIV-1) infection. *J Clin Immunol*. 1993;13(5):289-301.
48. Ho GT, Aird RE, Liu B, Boyapati RK, Kennedy NA, Dorward DA, et al. MDR1 deficiency impairs mitochondrial homeostasis and promotes intestinal inflammation. *Mucosal Immunol*. 2018;11(1):120-30.
49. Bar F, Bochmann W, Widok A, von Medem K, Pagel R, Hirose M, et al. Mitochondrial gene polymorphisms that protect mice from colitis. *Gastroenterology*. 2013;145(5):1055-63 e3.
50. Biasi F, Leonarduzzi G, Oteiza PI, Poli G. Inflammatory bowel disease: mechanisms, redox considerations, and therapeutic targets. *Antioxid Redox Signal*. 2013;19(14):1711-47.